

Divergent contributions of regulatory T cells to the pathogenesis of chronic hepatitis C

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Abbreviations: DCs, dendritic cells; FoxP3, transcription factor forkhead box P3; HCV, hepatitis C virus; IFN, interferon; IL, interleukin; iT_{reg} cell, inducible regulatory T cell; nT_{reg} cell, natural regulatory T cell; T_{eff} cells, effector T cells; TGF- β , transforming growth factor-beta

Hepatitis C virus, a small single-stranded RNA virus, is a major cause of chronic liver disease. Resolution of primary hepatitis C virus infections depends upon the vigorous responses of CD4⁺ and CD8⁺ T cells to multiple viral epitopes. Although such broad CD4⁺ and CD8⁺ T-cell responses are readily detected early during the course of infection regardless of clinical outcome, they are not maintained in individuals who develop chronic disease. Purportedly, a variety of factors contribute to the diminished T-cell responses observed in chronic, virus-infected patients including the induction of and biological suppression by CD4⁺FoxP3⁺ regulatory T cells. Indeed, a wealth of evidence suggests that regulatory T cells play diverse roles in the pathogenesis of chronic hepatitis C, impairing the effector T-cell response and viral clearance early during the course of infection and suppressing liver injury as the disease progresses. The factors that affect the generation and biological response of regulatory T cells in chronic, hepatitis C virus-infected patients are discussed.

Introduction

Hepatitis C virus (HCV, a small single-stranded RNA virus) is the cause of hepatitis C, the most common blood-borne infection in the world; an estimated 180 million people globally, ~3 million in the US, are chronically infected.^{1,2} The RNA genome consists of a large open reading frame that encodes an approximate 3,000 amino acid polypeptide precursor that is cleaved by viral and cellular proteases to yield three structural proteins [core, envelope 1 (E1) and E2] and seven non-structural proteins, which are involved in viral synthesis.³ HCV is primarily transmitted via contact with infected blood or bodily fluids. The risk of contracting hepatitis C from contaminated blood or blood products in the US is negligible due to mandatory and reliable blood screening.⁴ Despite a safe blood supply, however, isolated nosocomial/iatrogenic infections continue to occur, emphasizing the need for enforcement of universal precautions and proper sterilization of medical devices.⁴ Individuals at highest risk of infection in the

US today are injection drug users.⁵ In contrast, a large percentage of the population in many developing countries is infected owing to substandard infrastructure and technology required to provide a safe blood supply.⁶

The majority of HCV-infected patients develops chronic disease; only an approximate 20% resolve infection spontaneously.² A variety of factors such as age, sex, ethnicity, human immunodeficiency virus or hepatitis B co-infection, and host genetics (e.g., IL28B gene variants and HLA haplotype) affect spontaneous clearance of HCV.^{7–10} Standard treatment of unresolved infections, consisting of the combined administration of PEGylated interferon (IFN) and ribavirin over an extended period of 24 to 48 weeks, is often accompanied by significant side effects. Moreover, only 45–50% of treated patients infected with HCV genotype 1 (the primary etiologic agent of hepatitis C in the US) demonstrate a sustained virologic response following treatment withdrawal.¹¹ Additional treatment approaches that combine protease inhibitors, e.g., boceprevir and telaprevir, with ribavirin and PEGylated IFN result in sustained virologic responses in up to 80% of patients infected with HCV genotype 1, though the severity of accompanying side effects can also increase dramatically.^{12–14}

The vigorous responses of HLA class I- (CD8⁺) and class II-restricted (CD4⁺) T cells to numerous viral epitopes derived from both structural and nonstructural proteins are required for the resolution of primary HCV infections.¹⁵ Such broad responses are readily detected during the early course of infection despite clinical outcome, but subside in patients who later develop chronic disease.¹⁶ While patients who spontaneously clear infection continue to exhibit sustained, proliferative responses to a broad array of class I- and class II-restricted epitopes, chronically infected individuals respond to only a limited number.¹⁷ Purportedly, multiple factors contribute to the reduction in T-cell responses found in patients with chronic infections, notably, the induction of and biological suppression by CD4⁺FoxP3⁺ regulatory T_(reg) cells.^{18–22}

Regulatory T Cells

Background. CD4⁺ regulatory T_(reg) cells constitute one of the major mechanisms underlying immunological homeostasis and

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self-tolerance.²³ T_{reg} cells are characterized by the constitutive expression of the transcription factor forkhead box P3 (FoxP3) and interleukin (IL)-2 receptor α chain (CD25) on the cell surface. FoxP3 mutation and T_{reg} cell dysfunction result in fatal immunopathology and autoimmune disease in both mice and men.²⁴⁻²⁶ T_{reg} cells also moderate effector immune responses to infectious diseases that, if sustained at elevated levels, can lead to serious host tissue and organ damage.²⁷ Although a key factor in the maintenance of immune homeostasis, T_{reg} cells can also suppress effective immune responses to autologous tumors (e.g., hematological malignancies and metastatic melanoma) and promote persistent infections by a wide variety of microbial pathogens.²⁷⁻²⁹

T_{reg} cell phenotype. Two distinct T_{reg} cell subsets, classically distinguished by site of origin, are described in the literature. Natural (n) T_{reg} cells are generated by high-avidity selection in the thymus; inducible (i) T_{reg} cells, on the other hand, derive from conventional (CD4⁺CD25⁻FoxP3⁻) T cells in the periphery following stimulation.³⁰⁻³² n T_{reg} cells can induce “infectious tolerance” by converting conventional T cells into iT_{reg} cells via two primary methods: cytokine (IL-10, IL-35 or TGF- β)-dependent and dendritic cell (DC)-mediated, cytokine-independent mechanisms.^{33,34} Purportedly, n T_{reg} and iT_{reg} cells possess complementary immune functions: prevention of autoimmunity and maintenance of a non-inflammatory environment, respectively.³¹

Notably, no specific marker defines T_{reg} cells or differentiates n T_{reg} and iT_{reg} cell subsets. While FoxP3 expression is a common attribute of both subsets, conventional human T cells lacking immunosuppressive capacity can also express FoxP3 transiently following activation.³² Moreover, despite the near exclusive expression of CD25 by n T_{reg} cells in naïve mice, CD25 is expressed by a much more heterogeneous T-cell population in humans.³² Recent studies report the high level expression of neuropilin-1 on the surface of n T_{reg} , but not iT_{reg}, cells in mice, enabling differentiation and separation of these two subsets.^{35,36} Activated human FoxP3⁺ T_{reg} cells that express high suppressive activity are also distinguished by presence of glycoprotein A repetitions predominant (GARP, or LRRC32), a cell surface transmembrane protein that contains leucine-rich repeats.³⁷⁻⁴⁰ GARP mRNA is specifically expressed by CD4⁺CD25^{hi} T_{reg} cells, and is rapidly upregulated following T-cell receptor engagement.^{37,38} GARP anchors transforming growth factor (TGF)- β to the cell surface conferring increased suppressive activity and the ability to induce infectious tolerance.³⁹ Lastly, cell surface expression of ectonucleotidase, CD39, distinguishes activated, effector memory T_{reg} cells capable of abrogating DC maturation and T cell-dependent cytotoxicity.⁴¹

T_{reg} Cell Function

Contact-independent mechanisms. Activated T_{reg} cells are able to suppress the activity of a variety of immune cell types, i.e., both CD8⁺ and CD4⁺ T cells, NK cells, NKT cells, B cells, macrophages and DCs.⁴²⁻⁴⁶ Multiple mechanisms contribute to this suppressive activity although it is widely believed that n T_{reg} cell-mediated suppression is dependent upon direct, cell-cell

contact.⁴⁶ The synthesis of inhibitory cytokines constitutes a principal contact-independent mechanism by which T_{reg} cells in general suppress T_{eff} cell activity (Fig. 1). Both the soluble and membrane-bound forms of TGF- β , for example, play key roles in inducing and/or maintaining iT_{reg} and n T_{reg} cells, and in suppressing conventional effector T_{eff} cell activation.^{45,47,48} Similarly, IL-10 plays a critical role in suppressing CD4⁺ T_{eff} cell responses to a variety of pathogens used in animal models, as well as those that contribute to human disease.²⁷

The constitutive, high-level expression of CD25 (IL-2 receptor α chain) constitutes an additional contact-independent mechanism underlying T_{reg} cell-mediated suppression. T_{reg} cells produce relatively low levels of IL-2 and, as such, require an exogenous source of IL-2 in order to proliferate and survive.⁴⁹ As a consequence of the rapid consumption of IL-2 by T_{reg} cells, T_{eff} cell populations are deprived of the cytokine necessary for activation.⁴⁹ The cell surface expression of CD39 and CD73 ectonucleotidases constitutes another mechanism by which T_{reg} cells disrupt the metabolic activity of T_{eff} cells.⁵⁰ The activity expressed by these molecules abrogates the proinflammatory response of T_{eff} cells by rapidly degrading extracellular ATP released by neighboring, activated or damaged cells.⁵⁰ Additionally, adenosine generated as a byproduct of ATP degradation further suppresses T_{eff} cell function by binding A2A receptors expressed on the cell surface and inducing T cell anergy.⁵⁰⁻⁵²

Contact-dependent mechanisms. A number of contact-dependent mechanisms also facilitate the ability of T_{reg} cells to suppress T_{eff} cell function. For example, T_{reg} cells can exhibit cytotoxic activity and induce T_{eff} cell apoptosis dependent upon the production of granzyme A, granzyme B and perforin.^{48,53} In addition, cell-surface galectin-1 appears to contribute to the immunosuppressive activity of T_{reg} cells.⁵⁴ A member of a highly conserved family of β -galactosidase-binding proteins, galectin-1 inhibits proliferation and promotes apoptosis of activated T_{eff} cells.⁵⁴

Apart from regulating T_{eff} cell function directly, T_{reg} cells can inhibit the maturation and immunostimulatory activity of DC and, consequently, suppress the activity of conventional T cells indirectly.⁴⁸ T_{reg} cells can render DCs tolerogenic and limit their capacity to sensitize naïve T cells by a process termed trans-endocytosis whereby cytotoxic T lymphocyte-4 (CTLA-4) molecules expressed by T_{reg} cells capture, internalize, and degrade co-stimulatory molecules (i.e., CD80 and CD86) expressed on the surface of DCs.⁵⁵ Lymphocyte activation gene-3 (LAG-3 of CD223), a CD4 homolog capable of binding MHC class II molecules, further suppresses DC maturation and the expression of co-stimulatory molecules.^{56,57} Lastly, neuropilin-1 expressed by n T_{reg} cells inhibits the sensitization of naïve CD4⁺ T cells by promoting extended interaction between T_{reg} cells and immature DCs.⁵⁸ Undoubtedly, the suppressive functions of T_{reg} cells depend upon a combination of the mechanisms described.

T_{reg} Cells in Hepatitis C

The increased numbers of T_{reg} cells present in the liver and peripheral blood of chronic HCV-infected patients relative to patients

who spontaneously clear infection suggests that T_{reg} cells play a significant role in the pathogenesis of chronic hepatitis C.^{18,19,27,59-63} In this regard, $CD4^+CD25^+$ T cells obtained from the peripheral blood of patients infected with HCV suppressed virus-specific proliferation and $IFN-\gamma$ production by $CD4^+$ and $CD8^+$ T cell; depletion of $CD25^+$ cells including T_{regs} from the total peripheral blood mononuclear cell (PBMC) population enhanced the proliferation of cells that remained.^{18,63} Whether the increased number of T_{reg} cells found in the livers of chronically-infected patients represents an virus-specific T_{reg} cell response or arises as a non-specific consequence of chronic inflammation and disease is not entirely clear.

HCV encodes T_{reg} cell epitopes. Researchers have identified T_{reg} cell epitopes present in both the structural and non-structural proteins synthesized by HCV.^{20,59,60,64-66} Using HCV peptide-loaded MHC class II tetramers to label and quantify the cells, Ebimuna and coworkers reported HCV epitope-specific recognition by $FOXP3^+$ T_{reg} cells present in the peripheral blood of chronically infected patients.⁶⁴ Moreover, a number of studies now report an increase in $FOXP3^+CD25^{high}$ cells that resemble nT_{reg} cells and express an anergic cytokine profile following stimulation with HCV derived epitopes.^{59,61,64} Comparing FACS-sorted $CD4^+CD25^{high}$ T cells (nT_{reg} phenotype) derived from uninfected and HCV-infected individuals, however, one study found only minimal differences in global gene expression.⁶⁴ In contrast, other investigators reported an increase in HCV epitope-specific T_{reg} cells that resembled iT_{reg} cells phenotypically and exhibited both IL-10- and TGF- β -dependent suppressive activity.^{60,65} Regardless, a general consensus supports the heterogeneous nature of the expanded T_{reg} cell population found in chronic, HCV-infected patients (i.e., composed of both nT_{reg} and iT_{reg} cell subsets). The factors that contribute to this expansion are neither well understood nor characterized. Conceivably, the initial response of nT_{reg} cells to infection promotes the conversion of conventional T cells to iT_{reg} cells and subsequent suppression (i.e., infectious tolerance).³⁴

HCV T_{reg} cell epitopes and human homology. The expressed function of nT_{reg} cells is to suppress autoimmunity and the response to self antigens.²³⁻²⁶ Activation of T_{reg} cells subsequent to HCV infection suggests the existence of viral epitopes that exhibit homology to self and the ability to simulate these nT_{reg} cells and viral persistence.^{59,67-70} Indeed, Kanduc and colleagues reported that 3003 unique pentapeptides comprised the HCV polyprotein; of these, 2760 pentapeptides occurred a total of

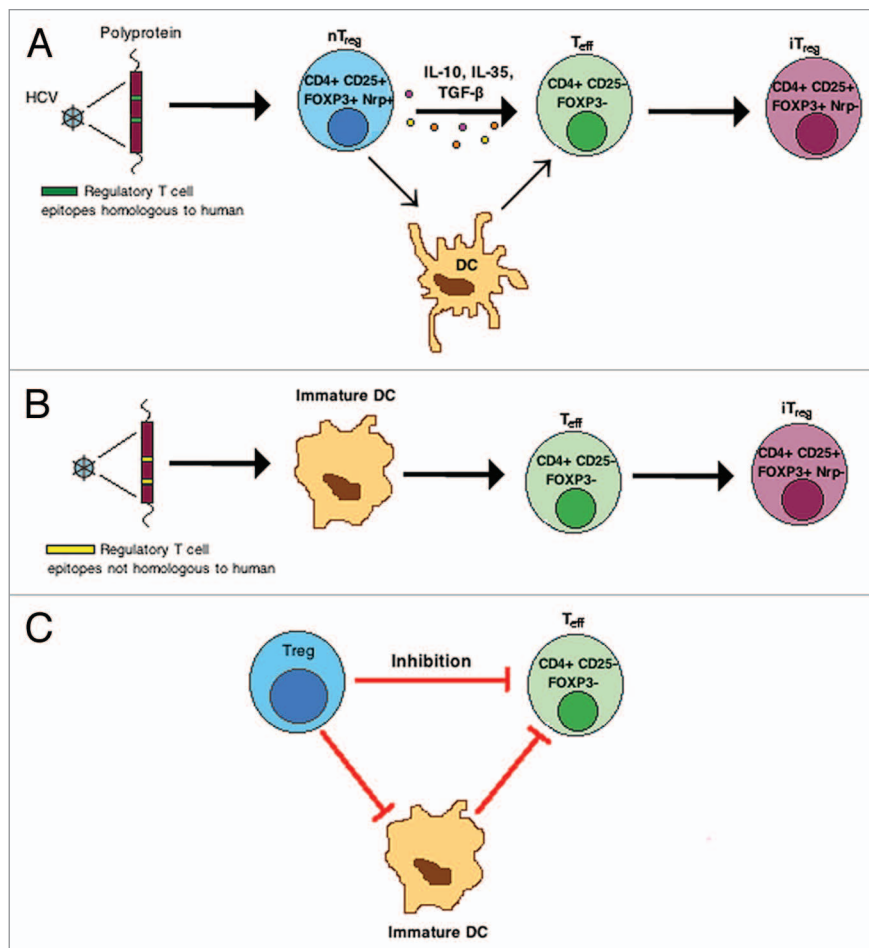
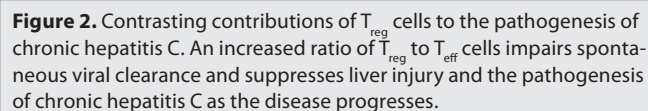


Figure 1. Increases in both the number and function of T_{reg} cells have been implicated in the pathogenesis of chronic hepatitis C. Virus-associated regulatory T cell epitopes, homologous to peptide sequences found in the human plasma proteome, induce nT_{reg} cell activation, conversion of T_{eff} to iT_{reg} cells and infectious tolerance (A). Viral epitopes lacking human homology, which are presented by immature DCs, elicit additional HCV-specific iT_{reg} cells (B). T_{reg} cells inhibit T_{eff} cell function by direct, contact-dependent and -independent mechanisms and by indirect mechanisms that affect DC maturation and/or immunostimulatory activity (C).

46,731 times (including matches/repeats) in 20,269 proteins that comprised the human proteome.^{68,70} Furthermore, a BLAST search and JanusMatrix analysis (a new computer algorithm, which identifies epitopes that are highly cross-reactive on the T cell receptor aspect) revealed extensive homology between a number of published T_{reg} cell epitopes derived from HCV structural and non-structural proteins and sequences encoded by the human genome (Moise et al., manuscript submitted) (Losikoff et al., manuscript in preparation).⁷¹ These epitopes induced the response of T_{reg} cells present in the peripheral blood of chronic HCV-infected patients, but not in blood obtained from HCV non-infected individuals.^{22,59,71} Taken together, these findings suggest that viral epitopes, homologous to peptide sequences constituting the human proteome, influence the pathogenesis of chronic hepatitis C by inducing the expansion of the iT_{reg} cell population and infectious tolerance.^{62,72}

An evolving body of research demonstrates the import of the human microbiome in shaping the immune response to viral



T_{reg} Cells Impair Viral Clearance

Recently, Cusick and coworkers described the emergence of a variant of an immunodominant HCV epitope during the course of chronic HCV infection.⁶⁶ This variant elicited a T_{reg} cell population that suppressed a T_{eff} cell response to the cognate, but not an unrelated, peptide. This finding suggests that variant, MHC class II-restricted viral epitopes arise as a consequence of immune pressure during the course of chronic HCV infection. Rather than induce the activity of conventional CD4⁺ T cells, these variant epitopes elicit an epitope-specific T_{reg} cell response that diminishes the CD4⁺ T cell help required for viral clearance. Thus, in cases of chronic disease, T_{reg} cells impair T_{eff}

T_{reg} Cells Suppress Chronic Hepatitis C-Associated Liver Injury

Thus, an equilibrium between T_{reg} cells and T_{eff} cells that permits both viral clearance and protection from HCV-related hepatopathology is critical for an optimal response to HCV infection.^{21,77} In cases of chronic disease, however, the elevated presence of T_{reg} cells appears to benefit both the pathogen (i.e., viral persistence) and the host (i.e., prevention of immunopathogenesis).^{21,79} In this regard, it is interesting to note that increased numbers of CD25⁺FoxP3⁺ T_{reg} cells were found in the livers of patients who underwent successful chemotherapy, suggesting that the continued presence of T_{reg} cells moderated immunopathology despite viral clearance. These divergent roles of T_{reg} cells during the development of chronic hepatitis C are illustrated in **Figure 2**.

Whether HCV infection elicits the response of T_{reg} cells directly or indirectly (dependent upon the intermediary role of another cell type) remains to be determined. Accumulating evidence suggests, however, that DCs play a key role in the expansion of T_{reg} cells during the pathogenesis of chronic disease. DCs are professional antigen presenting cells characterized by the potent capacity to elicit primary T cell responses.⁸¹ While there is no general consensus regarding the effects of HCV infection on DC function, it is often agreed that chronically infected patients

have significantly fewer DCs circulating in the peripheral blood, which correlates inversely with the severity of liver disease.⁸²⁻⁸⁶ Conversely, increased numbers of DCs are found in the liver of patients chronically infected with HCV.^{85,87-89} Regardless of their physiologic distribution (i.e., peripheral blood vs. liver), it is frequently suggested that the DCs are functionally impaired in patients with chronic hepatitis C.^{83,89-98} These impairments include lowered expression of HLA-DR and costimulatory molecules (e.g., CD86), decreased allostimulatory activity and the ability to secrete IFN- α and IL-12 and increased IL-10 production and the ability to prime T_{reg} cells.^{83-85,89,90,92-96,99-103} Additionally, it was reported that DCs derived from chronically infected patients remain phenotypically immature and lack the capacity to upregulate the expression of maturation or costimulatory marker following stimulation.^{91,97}

Mechanisms that underlie impaired DC function. The specific mechanisms underlying DC dysfunction in patients chronically infected with HCV remain to be clarified. Recent studies suggest, on the one hand, that DCs are susceptible to HCV infection.^{104,105} The interaction between envelope glycoproteins E1 and E2 and DC-SIGN, a C-type lectin expressed by DCs, mediates the ingestion of HCV viral particles.¹⁰⁵ This finding supports the speculation that, even in the absence of a productive infection, E1 and E2 bound to DC-SIGN transmit a signal that promotes T_{reg} cell formation and tolerance. Other structural and non-structural viral proteins further impair DC function by decreasing costimulatory molecule and HLA expression, reducing cytokine production, preventing TLR signaling and diminishing allostimulatory activity.¹⁰⁶ As described above, T_{reg} cells can also inhibit DC maturation and immunostimulatory capacity by a variety of mechanisms and, thereby, suppress the response of conventional T cells.⁴⁸ A growing body of evidence supports the critical role of DCs in the induction and maintenance of T_{reg} cells during chronic HCV infection, regardless of the specific mechanisms that contribute to a diminution in DC function.

DCs and immunological tolerance. DCs play a key role in establishing and maintaining immunological tolerance to both foreign and self antigens.^{107,108} Interactions between T_{reg} cells and DCs in the prototypic, tolerogenic environment maintained in the liver foster the development of chronic infectious diseases such as viral hepatitis.¹⁰⁹ While it is generally agreed that immature DCs promote T_{reg} cell function, the underlying mechanisms remain to be fully delineated.¹⁰⁷⁻¹¹¹ Notably, DCs obtained from human liver principally exhibit an immature phenotype characterized by low or negligible expression of cell-surface costimulatory molecule (e.g., CD40, CD80, CD83 and CD86).¹⁰⁹ Relative to DCs purified from the peripheral blood, DCs derived from liver also produce lower levels of proinflammatory cytokine (i.e., TNF- α , IL-1 β and IL-6) and significantly higher levels of IL-10 following stimulation. Moreover, the IL-10-dependent

production of CD4⁺CD25⁺FoxP3⁺ T_{reg} cells was greater in co-cultures that contained naïve, allogeneic CD4⁺ T cells and liver DCs, compared with co-cultures that contained DCs derived from blood.¹¹⁰ Similar results were obtained when allogeneic T cells and DCs obtained from chronic, HCV-infected patients were co-cultured, i.e., greater expansion of the T_{reg} cell population than observed in co-cultures that contained DCs obtained from healthy individuals.¹⁰³

Summary and Clinical Implications

Eighty to 85% of the patients in the US infected with HCV develop chronic disease.² Persistent viral infection correlates with increased T_{reg} cell number and function in the tissues of HCV-infected patients. While triple drug therapy (e.g., boceprevir or telaprevir administered in conjunction with interferon and ribavirin) increases the sustained virologic response, the adverse side effects are often severe, the costs are extraordinarily high, and a significant portion of treated patients remains infected.¹²⁻¹⁴ Additional approaches, e.g., therapeutic vaccination, are urgently needed. To date, four vaccine strategies have demonstrated varied and only limited success in clinical trials: recombinant protein, peptide, genetic or DNA-based and vector-mediated.^{112,113} Recent evidence suggests that viral epitopes may activate n T_{reg} cells and, subsequently, induce expansion of the i T_{reg} cell population and immunosuppression via the direct effects of T_{reg} cells on T_{eff} cell activity or by indirect effects dependent upon intervening DCs. DCs that exhibit an immature phenotype, in turn, can promote the expansion and activity of T_{reg} cells and, thus, play a key role in exacerbating the pathogenesis of chronic hepatitis C. As a consequence, future efforts to develop a therapeutic vaccine must employ strategies to avoid incorporating epitopes that promote T_{reg} cell activity.

In the absence of a therapeutic vaccine, a full understanding of the response of T_{reg} cells to HCV infection could lead to the development of treatments capable of sustaining T_{eff} cell-mediated immunity while limiting tissue damage.¹¹⁴ In this regard, continued research efforts are needed to determine the most effective means of manipulating the T_{reg} cell response in a clinical setting.¹⁸ Progress in understanding the role of T_{reg} cells and the factors that influence protective immunity to hepatitis C are hampered, however, by the lack of a good rodent model and the ability of HCV to infect humans and chimpanzees only.¹¹⁵

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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References

1. Alter MJ, Margolis HS, Krawczynski K, Judson FN, Mares A, Alexander WJ, et al. The natural history of community-acquired hepatitis C in the United States. The Sentinel Counties Chronic non-A, non-B Hepatitis Study Team. *N Engl J Med* 1992; 327:1899-905; PMID:1280771; <http://dx.doi.org/10.1056/NEJM199212313272702>
2. El-Serag HB. Hepatocellular carcinoma and hepatitis C in the United States. *Hepatology* 2002; 36(Suppl 1):S74-83; PMID:12407579; <http://dx.doi.org/10.1002/hep.1840360710>
3. Grakoui A, Wychowski C, Lin C, Feinstone SM, Rice CM. Expression and identification of hepatitis C virus polypeptide cleavage products. *J Virol* 1993; 67:1385-95; PMID:7679746
4. Healthcare-associated hepatitis B and C outbreaks reported to the Centers for Disease Control and Prevention (CDC) in 2008-2011. CDC Website 2012.
5. Amon JJ, Garfein RS, Ahdieh-Grant L, Armstrong GL, Ouellet LJ, Latka MH, et al. Prevalence of hepatitis C virus infection among injection drug users in the United States, 1994-2004. *Clin Infect Dis* 2008; 46:1852-8; PMID:18462109; <http://dx.doi.org/10.1086/588297>
6. Brown RS Jr., Gaglio PJ. Scope of worldwide hepatitis C problem. *Liver Transpl* 2003; 9:S10-3; PMID:14586889; <http://dx.doi.org/10.1053/jlts.2003.50244>
7. Jauncey M, Micallef JM, Gilmour S, Amin J, White PA, Rawlinson W, et al. Clearance of hepatitis C virus after newly acquired infection in injection drug users. *J Infect Dis* 2004; 190:1270-4; PMID:15346337; <http://dx.doi.org/10.1086/423943>
8. Rauch A, Gaudieri S, Thio C, Bocharon PY. Host genetic determinants of spontaneous hepatitis C clearance. *Pharmacogenomics* 2009; 10:1819-37; PMID:19891557; <http://dx.doi.org/10.2217/pgs.09.121>
9. Kuzushita N, Hayashi N, Moribe T, Katayama K, Kantō T, Nakatani S, et al. Influence of HLA haplotypes on the clinical courses of individuals infected with hepatitis C virus. *Hepatology* 1998; 27:240-4; PMID:9425943; <http://dx.doi.org/10.1002/hep.510270136>
10. Neumann-Haefelin C. Protective role of HLA-B27 in HIV and hepatitis C virus infection. *Dtsch Med Wochenschr* 2011; 136:320-4; PMID:21302207; <http://dx.doi.org/10.1055/s-0031-1272531>
11. Davis GL, Esteban-Mur R, Rustgi V, Hoefs J, Gordon SC, Trepo C, et al. International Hepatitis Interventional Therapy Group. Interferon alfa-2b alone or in combination with ribavirin for the treatment of relapse of chronic hepatitis C. *N Engl J Med* 1998; 339:1493-9; PMID:9819447; <http://dx.doi.org/10.1056/NEJM199811193392102>
12. Schaefer EA, Chung RT. Anti-hepatitis C virus drugs in development. *Gastroenterology* 2012; 142:1340-50, e1; PMID:22537441; <http://dx.doi.org/10.1053/j.gastro.2012.02.015>
13. US Food and Drug Administration. Incivek (telaprevir) in combination with drugs peginterferon alfa and ribavirin (incivek combination treatment): drug safety communication - serious skin reactions. Electronic Communication, USDA Website 2012.
14. Liu S, Cipriano LE, Holodniy M, Owens DK, Goldhaber-Fiebert JD. New protease inhibitors for the treatment of chronic hepatitis C: a cost-effectiveness analysis. *Ann Intern Med* 2012; 156:279-90; PMID:22351713; <http://dx.doi.org/10.7326/0003-4819-156-4-201202210-00005>
15. Lechner F, Wong DK, Dunbar PR, Chapman R, Chung RT, Dohrenwend B, et al. Analysis of successful immune responses in persons infected with hepatitis C virus. *J Exp Med* 2000; 191:1499-512; PMID:10790425; <http://dx.doi.org/10.1084/jem.191.9.1499>
16. Schulze Zur Wiesch J, Ciuffreda D, Lewis-Ximenez L, Kasprowski V, Nolan BE, Streeck H, et al. Broadly directed virus-specific CD4⁺ T cell responses are primed during acute hepatitis C infection, but rapidly disappear from human blood with viral persistence. *J Exp Med* 2012; 209:61-75; PMID:22213804; <http://dx.doi.org/10.1084/jem.20100388>
17. Day CL, Lauer GM, Robbins GK, McGovern B, Wurcel AG, Gandhi RT, et al. Broad specificity of virus-specific CD4⁺ T-helper-cell responses in resolved hepatitis C virus infection. *J Virol* 2002; 76:12584-95; PMID:12438584; <http://dx.doi.org/10.1128/JVI.76.24.12584-12595.2002>
18. Cabrera R, Tu Z, Xu Y, Firpi RJ, Rosen HR, Liu C, et al. An immunomodulatory role for CD4⁺CD25⁺ regulatory T lymphocytes in hepatitis C virus infection. *Hepatology* 2004; 40:1062-71; PMID:15486925; <http://dx.doi.org/10.1002/hep.20454>
19. Ward SM, Fox BC, Brown PJ, Worthington J, Fox SB, Chapman RW, et al. Quantification and localisation of FOXP3⁺ T lymphocytes and relation to hepatic inflammation during chronic HCV infection. *J Hepatol* 2007; 47:316-24; PMID:17475362; <http://dx.doi.org/10.1016/j.jhep.2007.03.023>
20. Li S, Floess S, Hamann A, Gaudieri S, Lucas A, Hellard M, et al. Analysis of FOXP3⁺ regulatory T cells that display apparent viral antigen specificity during chronic hepatitis C virus infection. *PLoS Pathog* 2009; 5:e1000707; PMID:20041222; <http://dx.doi.org/10.1371/journal.ppat.1000707>
21. Sturm N, Thélus M, Camous X, Dimitrov G, Ramzan M, Dufeu-Duchesne T, et al. Characterization and role of intra-hepatic regulatory T cells in chronic hepatitis C pathogenesis. *J Hepatol* 2010; 53:25-35; PMID:20452085; <http://dx.doi.org/10.1016/j.jhep.2010.02.024>
22. Li S, Gowans EJ, Choungnet C, Plebanski M, Dittmer U. Natural regulatory T cells and persistent viral infection. *J Virol* 2008; 82:21-30; PMID:1785537; <http://dx.doi.org/10.1128/JVI.01768-07>
23. Sakaguchi S, Miyara M, Costantino CM, Hafler DA. FOXP3⁺ regulatory T cells in the human immune system. *Nat Rev Immunol* 2010; 10:490-500; PMID:20559327; <http://dx.doi.org/10.1038/nri2785>
24. Shull MM, Ormsby I, Kier AB, Pawlowski S, Diebold RJ, Yin M, et al. Targeted disruption of the mouse transforming growth factor- β gene results in multifocal inflammatory disease. *Nature* 1992; 359:693-9; PMID:1436033; <http://dx.doi.org/10.1038/359693a0>
25. Bennett CL, Christie J, Ramsdell F, Brunkow ME, Ferguson PJ, Whitesell L, et al. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat Genet* 2001; 27:20-1; PMID:11137993; <http://dx.doi.org/10.1038/83713>
26. Khattri R, Cox T, Yasayko SA, Ramsdell F. An essential role for Scurfin in CD4⁺CD25⁺ T regulatory cells. *Nat Immunol* 2003; 4:337-42; PMID:12612581; <http://dx.doi.org/10.1038/ni909>
27. Belkaid Y, Tarbell K. Regulatory T cells in the control of host-microorganism interactions (*). *Annu Rev Immunol* 2009; 27:551-89; PMID:19302048; <http://dx.doi.org/10.1146/annurev.immunol.021908.132723>
28. Tang Q, Bluestone JA. The Foxp3⁺ regulatory T cell: a jack of all trades, master of regulation. *Nat Immunol* 2008; 9:239-44; PMID:18285775; <http://dx.doi.org/10.1038/ni1572>
29. Cao X, Cai SF, Fehniger TA, Song J, Collins LI, Piwnicka-Worms DR, et al. Granzyme B and perforin are important for regulatory T cell-mediated suppression of tumor clearance. *Immunity* 2007; 27:635-46; PMID:17919943; <http://dx.doi.org/10.1016/j.immuni.2007.08.014>
30. Bluestone JA, Abbas AK. Natural versus adaptive regulatory T cells. *Nat Rev Immunol* 2003; 3:253-7; PMID:12658273; <http://dx.doi.org/10.1038/nri1032>
31. Curotto de Lafaille MA, Lafaille JJ. Natural and adaptive foxp3⁺ regulatory T cells: more of the same or a division of labor? *Immunity* 2009; 30:626-35; PMID:19464985; <http://dx.doi.org/10.1016/j.immuni.2009.05.002>
32. Miyara M, Sakaguchi S. Human Foxp3⁺CD4⁺ regulatory T cells: their knowns and unknowns. *Immunol Cell Biol* 2011; 89:346-51; PMID:21301480; <http://dx.doi.org/10.1038/icb.2010.137>
33. Jonuleit H, Schmitt E, Kakirman H, Stassen M, Knop J, Enk AH. Infectious tolerance: human CD25⁺ regulatory T cells convey suppressor activity to conventional CD4⁺ T helper cells. *J Exp Med* 2002; 196:255-60; PMID:12119350; <http://dx.doi.org/10.1084/jem.20020394>
34. Gravano DM, Vignali DA. The battle against immunopathology: infectious tolerance mediated by regulatory T cells. *Cell Mol Life Sci* 2012; 69:1997-2008; PMID:22205213; <http://dx.doi.org/10.1007/s00018-011-0907-z>
35. Yadav M, Louvet C, Davini D, Gardner JM, Martínez-Llordella M, Bailey-Bucktrout S, et al. Neuropilin-1 distinguishes natural and inducible regulatory T cells among regulatory T cell subsets in vivo. *J Exp Med* 2012; 209:1713-22, S1-19; PMID:22966003; <http://dx.doi.org/10.1084/jem.20120822>
36. Weiss JM, Bilate AM, Gobert M, Ding Y, Curotto de Lafaille M, Parkhurst CN, et al. Neuropilin 1 is expressed on thymus-derived natural regulatory T cells, but not mucosa-generated induced Foxp3⁺ T reg cells. *J Exp Med* 2012; 209:1723-42, S1.
37. Wang R, Wan Q, Kozhaya L, Fujii H, Unutmaz D. Identification of a regulatory T cell specific cell surface molecule that mediates suppressive signals and induces Foxp3 expression. *PLoS One* 2008; 3:e2705; PMID:18628982; <http://dx.doi.org/10.1371/journal.pone.0002705>
38. Wang R, Kozhaya L, Mercer F, Khaitan A, Fujii H, Unutmaz D. Expression of GARP selectively identifies activated human FOXP3⁺ regulatory T cells. *Proc Natl Acad Sci U S A* 2009; 106:13439-44; PMID:19666573
39. Tran DQ, Andersson J, Wang R, Ramsey H, Unutmaz D, Shevach EM. GARP (LRRC32) is essential for the surface expression of latent TGF-beta on platelets and activated FOXP3⁺ regulatory T cells. *Proc Natl Acad Sci U S A* 2009; 106:13445-50; PMID:19651619; <http://dx.doi.org/10.1073/pnas.0901944106>
40. Stockis J, Colau D, Coulie PG, Lucas S. Membrane protein GARP is a receptor for latent TGF-beta on the surface of activated human Treg. *Eur J Immunol* 2009; 39:3315-22; PMID:19750484; <http://dx.doi.org/10.1002/eji.200939684>
41. Dwyer KM, Hanidziar D, Putheti P, Hill PA, Pommey S, McRae JL, et al. Expression of CD39 by human peripheral blood CD4⁺ CD25⁺ T cells denotes a regulatory memory phenotype. *Am J Transplant* 2010; 10:2410-20; PMID:20977632; <http://dx.doi.org/10.1111/j.1600-6143.2010.03291.x>
42. Zhao DM, Thornton AM, DiPaolo RJ, Shevach EM. Activated CD4⁺CD25⁺ T cells selectively kill B lymphocytes. *Blood* 2006; 107:3925-32; PMID:16418326; <http://dx.doi.org/10.1182/blood-2005-11-4502>
43. Ghiringhelli F, Ménard C, Martin F, Zitvogel L. The role of regulatory T cells in the control of natural killer cells: relevance during tumor progression. *Immunol Rev* 2006; 214:229-38; PMID:17100888; <http://dx.doi.org/10.1111/j.1600-065X.2006.00445.x>
44. La Cava A, Van Kaer L, Fu-Dong-Shi. CD4⁺CD25⁺ Tregs and NKT cells: regulators regulating regulators. *Trends Immunol* 2006; 27:322-7; PMID:16735139; <http://dx.doi.org/10.1016/j.it.2006.05.003>
45. Shevach EM. Mechanisms of foxp3⁺ T regulatory cell-mediated suppression. *Immunity* 2009; 30:636-45; PMID:19464986; <http://dx.doi.org/10.1016/j.immuni.2009.04.010>

46. Miyara M, Sakaguchi S. Natural regulatory T cells: mechanisms of suppression. *Trends Mol Med* 2007; 13:108-16; PMID:17257897; <http://dx.doi.org/10.1016/j.molmed.2007.01.003>
47. Andersson J, Tran DQ, Pesu M, Davidson TS, Ramsey H, O'Shea JJ, et al. CD4⁺ FoxP3⁺ regulatory T cells confer infectious tolerance in a TGF-beta-dependent manner. *J Exp Med* 2008; 205:1975-81; PMID:18710931; <http://dx.doi.org/10.1084/jem.20080308>
48. Vignali DA, Collison LW, Workman CJ. How regulatory T cells work. *Nat Rev Immunol* 2008; 8:523-32; PMID:18566595; <http://dx.doi.org/10.1038/nri2343>
49. Yamaguchi T, Wing JB, Sakaguchi S. Two modes of immune suppression by Foxp3⁺ regulatory T cells under inflammatory or non-inflammatory conditions. *Semin Immunol* 2011; 23:424-30; PMID:22055883; <http://dx.doi.org/10.1016/j.smim.2011.10.002>
50. Borsellino G, Kleinschewitz M, Di Mitri D, Sternjak A, Diamantini A, Giometto R, et al. Expression of ectonucleotidase CD39 by Foxp3⁺ Treg cells: hydrolysis of extracellular ATP and immune suppression. *Blood* 2007; 110:1225-32; PMID:17449799; <http://dx.doi.org/10.1182/blood-2006-12-064527>
51. Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, et al. Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. *J Exp Med* 2007; 204:1257-65; PMID:17502665; <http://dx.doi.org/10.1084/jem.20062512>
52. Yip L, Woehrle T, Corriden R, Hirsh M, Chen Y, Inoue Y, et al. Autocrine regulation of T-cell activation by ATP release and P2X7 receptors. *FASEB J* 2009; 23:1685-93; PMID:19211924; <http://dx.doi.org/10.1096/fj.08-126458>
53. Ren X, Ye F, Jiang Z, Chu Y, Xiong S, Wang Y. Involvement of cellular death in TRAIL/DR5-dependent suppression induced by CD4⁺CD25⁺ regulatory T cells. *Cell Death Differ* 2007; 14:2076-84; PMID:17762882; <http://dx.doi.org/10.1038/sj.cdd.4402220>
54. Garín MI, Chu CC, Golshayan D, Cernuda-Morollón E, Wait R, Lechler RI. Galectin-1: a key effector of regulation mediated by CD4⁺CD25⁺ T cells. *Blood* 2007; 109:2058-65; PMID:17110462; <http://dx.doi.org/10.1182/blood-2006-04-016451>
55. Qureshi OS, Zheng Y, Nakamura K, Attridge K, Manzotti C, Schmidt EM, et al. Trans-endocytosis of CD80 and CD86: a molecular basis for the cell-extrinsic function of CTLA-4. *Science* 2011; 332:600-3; PMID:21474713; <http://dx.doi.org/10.1126/science.1202947>
56. Huang CT, Workman CJ, Flies D, Pan X, Marson AL, Zhou G, et al. Role of LAG-3 in regulatory T cells. *Immunity* 2004; 21:503-13; PMID:15485628; <http://dx.doi.org/10.1016/j.immuni.2004.08.010>
57. Liang B, Workman C, Lee J, Chew C, Dale BM, Colonna L, et al. Regulatory T cells inhibit dendritic cells by lymphocyte activation gene-3 engagement of MHC class II. *J Immunol* 2008; 180:5916-26; PMID:18424711
58. Sarris M, Andersen KG, Randow F, Mayr L, Betz AG. Neuropilin-1 expression on regulatory T cells enhances their interactions with dendritic cells during antigen recognition. *Immunity* 2008; 28:402-13; PMID:18328743; <http://dx.doi.org/10.1016/j.immuni.2008.01.012>
59. Li S, Jones KL, Woollard DJ, Dromey J, Paukovics G, Plebanski M, et al. Defining target antigens for CD25⁺ FOXP3⁺ IFN-γ⁺ regulatory T cells in chronic hepatitis C virus infection. *Immunol Cell Biol* 2007; 85:197-204; PMID:17199111
60. MacDonald AJ, Duffy M, Brady MT, McKiernan S, Hall W, Hegarty J, et al. CD4⁺ T helper type 1 and regulatory T cells induced against the same epitopes on the core protein in hepatitis C virus-infected persons. *J Infect Dis* 2002; 185:720-7; PMID:11920289; <http://dx.doi.org/10.1086/339340>
61. Smyk-Pearson S, Golden-Mason L, Klarquist J, Burton JR Jr., Tester IA, Wang CC, et al. Functional suppression by FoxP3⁺CD4⁺CD25⁺(high) regulatory T cells during acute hepatitis C virus infection. *J Infect Dis* 2008; 197:46-57; PMID:18171284; <http://dx.doi.org/10.1086/523651>
62. Boettler T, Spangenberg HC, Neumann-Haefelin C, Panther E, Urbani S, Ferrari C, et al. T cells with a CD4⁺CD25⁺ regulatory phenotype suppress *in vitro* proliferation of virus-specific CD8⁺ T cells during chronic hepatitis C virus infection. *J Virol* 2005; 79:7860-7; PMID:15919940; <http://dx.doi.org/10.1128/JVI.79.12.7860-7867.2005>
63. Sugimoto K, Ikeda F, Stadanlick J, Nunes FA, Alter HJ, Chang KM. Suppression of HCV-specific T cells without differential hierarchy demonstrated *ex vivo* in persistent HCV infection. *Hepatology* 2003; 38:1437-48; PMID:14647055
64. Ebinuma H, Nakamoto N, Li Y, Price DA, Gostick E, Levine BL, et al. Identification and *in vitro* expansion of functional antigen-specific CD25⁺ FoxP3⁺ regulatory T cells in hepatitis C virus infection. *J Virol* 2008; 82:5043-53; PMID:18337568; <http://dx.doi.org/10.1128/JVI.01548-07>
65. Langhans B, Braunschweiger I, Arndt S, Schulte W, Satoguina J, Layland LE, et al. Core-specific adaptive regulatory T-cells in different outcomes of hepatitis C. *Clin Sci (Lond)* 2010; 119:97-109; PMID:20222873; <http://dx.doi.org/10.1042/CS20090661>
66. Cusick MF, Schiller JJ, Gill JC, Eckels DD. Hepatitis C virus induces regulatory T cells by naturally occurring viral variants to suppress T cell responses. *Clin Dev Immunol* 2011; 2011:806061.
67. Mishiro S, Takeda K, Hoshi Y, Yoshikawa A, Gotanda T, Itoh Y. An autoantibody cross-reactive to hepatitis C virus core and a host nuclear antigen. *Autoimmunity* 1991; 10:269-73; PMID:1723001; <http://dx.doi.org/10.3109/08916939109001900>
68. Kanduc D, Stufano A, Lucchese G, Kusalik A. Massive peptide sharing between viral and human proteomes. *Peptides* 2008; 29:1755-66; PMID:18582510; <http://dx.doi.org/10.1016/j.peptides.2008.05.022>
69. Kanduc D. HCV: Written in our DNA. *Self Nonself* 2011; 2:108-13; PMID:22299062; <http://dx.doi.org/10.4161/self.2.2.15795>
70. Kusalik A, Bickis M, Lewis C, Li Y, Lucchese G, Marincola FM, et al. Widespread and ample peptide overlapping between HCV and Homo sapiens proteomes. *Peptides* 2007; 28:1260-7; PMID:17485143; <http://dx.doi.org/10.1016/j.peptides.2007.04.001>
71. Losikoff P, Terry F, Mishra S, Martin W, Ardito M, Bailey-Kellogg C, et al. A hepatitis C virus-encoded epitope, homologous to a wide array of predicted human epitopes, stimulates a T-regulatory cell response in patients with chronic hepatitis C. *New England Regional Center of Excellence Annual Retreat* 2012 2012.
72. Rushbrook SM, Ward SM, Unitt E, Vowler SL, Lucas M, Klennerman P, et al. Regulatory T cells suppress *in vitro* proliferation of virus-specific CD8⁺ T cells during persistent hepatitis C virus infection. *J Virol* 2005; 79:7852-9; PMID:15919939; <http://dx.doi.org/10.1128/JVI.79.12.7852-7859.2005>
73. Pfeiffer JK, Sonnenburg JL. The intestinal microbiota and viral susceptibility. *Front Microbiol* 2011; 2:92; PMID:21833331; <http://dx.doi.org/10.3389/fmicb.2011.00092>
74. Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, et al. Induction of colonic regulatory T cells by indigenous Clostridium species. *Science* 2011; 331:337-41; PMID:21205640; <http://dx.doi.org/10.1126/science.1198469>
75. Lathrop SK, Bloom SM, Rao SM, Nutsch K, Lio CW, Santacruz N, et al. Peripheral education of the immune system by colonic commensal microbiota. *Nature* 2011; 478:250-4; PMID:21937990; <http://dx.doi.org/10.1038/nature10434>
76. Heeg MH, Ulsenheimer A, Grüner NH, Zachoval R, Jung MC, Gerlach JT, et al. FOXP3 expression in hepatitis C virus-specific CD4⁺ T cells during acute hepatitis C. *Gastroenterology* 2009; 137:1280-8; e1-6; PMID:19596013; <http://dx.doi.org/10.1053/j.gastro.2009.06.059>
77. Manigold T, Racanelli V. T-cell regulation by CD4 regulatory T cells during hepatitis B and C virus infections: facts and controversies. *Lancet Infect Dis* 2007; 7:804-13; PMID:18045563; [http://dx.doi.org/10.1016/S1473-3099\(07\)70289-X](http://dx.doi.org/10.1016/S1473-3099(07)70289-X)
78. Speletas M, Argentou N, Germanidis G, Vasiladis T, Mantzoukis K, Patsiaoura K, et al. Foxp3 expression in liver correlates with the degree but not the cause of inflammation. *Mediators Inflamm* 2011; 2011:827565; PMID:21772667; <http://dx.doi.org/10.1155/2011/827565>
79. Rouse BT, Suvas S. Regulatory cells and infectious agents: detentes cordiales et contraire. *J Immunol* 2004; 173:2211-5; PMID:15294929
80. Bolacchi F, Sinistro A, Ciaprin C, Demin F, Capozzi M, Carducci FC, et al. Increased hepatitis C virus (HCV)-specific CD4⁺CD25⁺ regulatory T lymphocytes and reduced HCV-specific CD4⁺ T cell response in HCV-infected patients with normal versus abnormal alanine aminotransferase levels. *Clin Exp Immunol* 2006; 144:188-96; PMID:16634790; <http://dx.doi.org/10.1111/j.1365-2249.2006.03048.x>
81. Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature* 1998; 392:245-52; PMID:9521319; <http://dx.doi.org/10.1038/32588>
82. Ryan EJ, O'Farrelly C. The effect of chronic hepatitis C infection on dendritic cell function: a summary of the experimental evidence. *J Viral Hepat* 2011; 18:601-7; PMID:21794024; <http://dx.doi.org/10.1111/j.1365-2893.2011.01453.x>
83. Ulsenheimer A, Gerlach JT, Jung MC, Gruener N, Wächter M, Backmund M, et al. Plasmacytoid dendritic cells in acute and chronic hepatitis C virus infection. *Hepatology* 2005; 41:643-51; PMID:15726647; <http://dx.doi.org/10.1002/hep.20592>
84. Rehmann B. Hepatitis C virus versus innate and adaptive immune responses: a tale of coevolution and coexistence. *J Clin Invest* 2009; 119:1745-54; PMID:19587449; <http://dx.doi.org/10.1172/JCI39133>
85. Cicinnati VR, Kang J, Sotiropoulos GC, Hilgard P, Frilling A, Broelsch CE, et al. Altered chemotactic response of myeloid and plasmacytoid dendritic cells from patients with chronic hepatitis C: role of alpha interferon. *J Gen Virol* 2008; 89:1243-53; PMID:18420803; <http://dx.doi.org/10.1099/vir.0.83517-0>
86. Kunitani H, Shimizu Y, Murata H, Higuchi K, Watanabe A. Phenotypic analysis of circulating and intrahepatic dendritic cell subsets in patients with chronic liver diseases. *J Hepatol* 2002; 36:734-41; PMID:12044522; [http://dx.doi.org/10.1016/S0168-8278\(02\)00062-4](http://dx.doi.org/10.1016/S0168-8278(02)00062-4)
87. Nattermann J, Zimmermann H, Iwan A, von Lilienfeld-Toal M, Leifeld L, Nischalke HD, et al. Hepatitis C virus E2 and CD81 interaction may be associated with altered trafficking of dendritic cells in chronic hepatitis C. *Hepatology* 2006; 44:945-54; PMID:17006905; <http://dx.doi.org/10.1002/hep.21350>
88. Wald O, Weiss ID, Galun E, Peled A. Chemokines in hepatitis C virus infection: pathogenesis, prognosis and therapeutics. *Cytokine* 2007; 39:50-62; PMID:17629707; <http://dx.doi.org/10.1016/j.cyt.2007.05.013>
89. Wertheimer AM, Bakke A, Rosen HR. Direct enumeration and functional assessment of circulating dendritic cells in patients with liver disease. *Hepatology* 2004; 40:335-45; PMID:15368438; <http://dx.doi.org/10.1002/hep.20306>

90. Kanto T, Hayashi N, Takehara T, Tatsumi T, Kuzushita N, Ito A, et al. Impaired allostimulatory capacity of peripheral blood dendritic cells recovered from hepatitis C virus-infected individuals. *J Immunol* 1999; 162:5584-91; PMID:10228041
91. Auffermann-Gretzinger S, Keeffe EB, Levy S. Impaired dendritic cell maturation in patients with chronic, but not resolved, hepatitis C virus infection. *Blood* 2001; 97:3171-6; PMID:11342445; <http://dx.doi.org/10.1182/blood.V97.10.3171>
92. Anthony DD, Yonkers NL, Post AB, Asaad R, Heinzel FP, Lederman MM, et al. Selective impairments in dendritic cell-associated function distinguish hepatitis C virus and HIV infection. *J Immunol* 2004; 172:4907-16; PMID:15067070
93. Murakami H, Akbar SM, Matsui H, Horiike N, Onji M. Decreased interferon-alpha production and impaired T helper 1 polarization by dendritic cells from patients with chronic hepatitis C. *Clin Exp Immunol* 2004; 137:559-65; PMID:15320906; <http://dx.doi.org/10.1111/j.1365-2249.2004.02550.x>
94. Szabo G, Dolganiuc A. Subversion of plasmacytoid and myeloid dendritic cell functions in chronic HCV infection. *Immunobiology* 2005; 210:237-47; PMID:16164031; <http://dx.doi.org/10.1016/j.imbio.2005.05.018>
95. Della Bella S, Crocignani A, Riva A, Presicce P, Benetti A, Longhi R, et al. Decrease and dysfunction of dendritic cells correlate with impaired hepatitis C virus-specific CD4⁺ T-cell proliferation in patients with hepatitis C virus infection. *Immunology* 2007; 121:283-92; PMID:17462079; <http://dx.doi.org/10.1111/j.1365-2567.2007.02577.x>
96. Averill L, Lee WM, Karandikar NJ. Differential dysfunction in dendritic cell subsets during chronic HCV infection. *Clin Immunol* 2007; 123:40-9; PMID:17239662; <http://dx.doi.org/10.1016/j.clim.2006.12.001>
97. Gelderblom HC, Nijhuis LE, de Jong EC, te Velde AA, Pajkrt D, Reesink HW, et al. Monocyte-derived dendritic cells from chronic HCV patients are not infected but show an immature phenotype and aberrant cytokine profile. *Liver Int* 2007; 27:944-53; PMID:17696933; <http://dx.doi.org/10.1111/j.1478-3231.2007.01507.x>
98. MacDonald AJ, Semper AE, Libri NA, Rosenberg WM. Monocyte-derived dendritic cell function in chronic hepatitis C is impaired at physiological numbers of dendritic cells. *Clin Exp Immunol* 2007; 148:494-500; PMID:17362265; <http://dx.doi.org/10.1111/j.1365-2249.2007.03367.x>
99. Kanto T, Inoue M, Miyatake H, Sato A, Sakakibara M, Yakushiji T, et al. Reduced numbers and impaired ability of myeloid and plasmacytoid dendritic cells to polarize T helper cells in chronic hepatitis C virus infection. *J Infect Dis* 2004; 190:1919-26; PMID:15529255; <http://dx.doi.org/10.1086/425425>
100. Saito K, Ait-Goughoulte M, Truscott SM, Meyer K, Blazevic A, Abate G, et al. Hepatitis C virus inhibits cell surface expression of HLA-DR, prevents dendritic cell maturation, and induces interleukin-10 production. *J Virol* 2008; 82:3320-8; PMID:18216090; <http://dx.doi.org/10.1128/JVI.02547-07>
101. Mengshol JA, Golden-Mason L, Castelblanco N, Im KA, Dillon SM, Wilson CC, et al.; Virahep-C Study Group. Impaired plasmacytoid dendritic cell maturation and differential chemotaxis in chronic hepatitis C virus: associations with antiviral treatment outcomes. *Gut* 2009; 58:964-73; PMID:19193669; <http://dx.doi.org/10.1136/gut.2008.168948>
102. Bain C, Fatmi A, Zoulim F, Zarski JP, Trépo C, Inchauspé G. Impaired allostimulatory function of dendritic cells in chronic hepatitis C infection. *Gastroenterology* 2001; 120:512-24; PMID:11159892; <http://dx.doi.org/10.1053/gast.2001.21212>
103. Dolganiuc A, Paek E, Kodys K, Thomas J, Szabo G. Myeloid dendritic cells of patients with chronic HCV infection induce proliferation of regulatory T lymphocytes. *Gastroenterology* 2008; 135:2119-27; PMID:18835391; <http://dx.doi.org/10.1053/j.gastro.2008.07.082>
104. Blackard JT, Smeaton L, Hiasa Y, Horiike N, Onji M, Jamieson DJ, et al. Detection of hepatitis C virus (HCV) in serum and peripheral-blood mononuclear cells from HCV-monoinfected and HIV/HCV-coinfected persons. *J Infect Dis* 2005; 192:258-65; PMID:15962220; <http://dx.doi.org/10.1086/430949>
105. Ludwig IS, Lekkerkerker AN, Depla E, Bosman F, Musters RJ, Depraetere S, et al. Hepatitis C virus targets DC-SIGN and L-SIGN to escape lysosomal degradation. *J Virol* 2004; 78:8322-32; PMID:15254204; <http://dx.doi.org/10.1128/JVI.78.15.8322-8332.2004>
106. Krishnadas DK, Ahn JS, Han J, Kumar R, Agrawal B. Immunomodulation by hepatitis C virus-derived proteins: targeting human dendritic cells by multiple mechanisms. *Int Immunol* 2010; 22:491-502; PMID:20410260; <http://dx.doi.org/10.1093/intimm/dxq033>
107. Steinman RM, Hawiger D, Nussenzweig MC. Tolerogenic dendritic cells. *Annu Rev Immunol* 2003; 21:685-711; PMID:12615891; <http://dx.doi.org/10.1146/annurev.immunol.21.120601.141040>
108. Maldonado RA, von Andrian UH. How tolerogenic dendritic cells induce regulatory T cells. *Adv Immunol* 2010; 108:111-65; PMID:21056730; <http://dx.doi.org/10.1016/B978-0-12-380995-7.00004-5>
109. Tiegs G, Lohse AW. Immune tolerance: what is unique about the liver. *J Autoimmun* 2010; 34:1-6; PMID:19717280; <http://dx.doi.org/10.1016/j.jaut.2009.08.008>
110. Bamboat ZM, Stableford JA, Plitas G, Burt BM, Nguyen HM, Welles AP, et al. Human liver dendritic cells promote T cell hyporesponsiveness. *J Immunol* 2009; 182:1901-11; PMID:19201843; <http://dx.doi.org/10.4049/jimmunol.0803404>
111. Cobbold SP, Adams E, Nolan KE, Regateiro FS, Waldmann H. Connecting the mechanisms of T-cell regulation: dendritic cells as the missing link. *Immunol Rev* 2010; 236:203-18; PMID:20636819; <http://dx.doi.org/10.1111/j.1600-065X.2010.00913.x>
112. Yu CI, Chiang BL. A new insight into hepatitis C vaccine development. *J Biomed Biotechnol* 2010; 2010:548280; PMID:20625493; <http://dx.doi.org/10.1155/2010/548280>
113. Halliday J, Klenerman P, Barnes E. Vaccination for hepatitis C virus: closing in on an evasive target. *Expert Rev Vaccines* 2011; 10:659-72; PMID:21604986; <http://dx.doi.org/10.1586/erv.11.55>
114. Keynan Y, Card CM, McLaren PJ, Dawood MR, Kasper K, Fowke KR. The role of regulatory T cells in chronic and acute viral infections. *Clin Infect Dis* 2008; 46:1046-52; PMID:18444822; <http://dx.doi.org/10.1086/529379>
115. Billerbeck E, de Jong Y, Dorner M, de la Fuente C, Ploss A. Animal models for hepatitis C. *Curr Top Microbiol Immunol* 2013; 369:49-86; PMID:23463197; http://dx.doi.org/10.1007/978-3-642-27340-7_3